

Evaluation of TNF- α , IL-8, and IL-6 in Thyroid Disorder Patients Leading to Thyroidectomy

Mohammed Shwish SALEH¹, Mohammad Ahmad ABDALLA^{2,*}, Mousa Jasim MOHAMMED¹

Abstract

Introduction: This study was carried out to investigate the immunological role of some cytokines in patients suffering from thyroid disorders (nodular hyperplasia, thyroid cancer, hypothyroidism, and thyrotoxicosis). Therefore, this study aimed to assess several immunological factors associated with clinically diagnosed thyroid disorders in patients who decided to undergo thyroidectomy.

Methods: It was conducted at specialized outpatient and governorate laboratories in the period from September 2024 to November 2025. Eighty female individuals participated in this study, and serum samples were collected from these individuals before the operation. Alongside them, another set of serum samples was collected from twenty healthy female individuals to compare different cytokine levels between these two groups. ELISA kits were used to evaluate the serum levels of tumor necrosis factor α (TNF- α), interleukin 8 (IL-8), and interleukin 6 (IL-6).

Results: In non-toxic goiter patients, there was a significant increase ($P \leq 0.01$) in the IL-8 and IL-6 levels; there were no significant differences between non-toxic goiter patients and the control group in the levels of TNF- α . In toxic goiter patients, there was a significant increase ($P \leq 0.01$) in the TNF- α , IL-8, and IL-6 levels. In hypothyroidism patients, there was a significant increase ($P \leq 0.01$) in the TNF- α , IL-8, and IL-6 levels. Also, in thyroid cancer patients, there was a significant increase ($P \leq 0.01$) in the TNF- α , IL-8, and IL-6 levels.

Conclusion: The present study results support the fact that cytokines play essential roles in the development and prognosis of thyroid disorders. It indicates that measurements (TNF- α , IL-8, and IL-6) can be considered diagnostic tests to predict thyroid disorders.

Keywords: Goiter, thyroid gland, tumor necrosis factor α , interleukin 8, interleukin 6.

¹Tikrit University - College of Sciences, Department of Biology, Tikrit, Iraq.

²Tikrit University - College of Medicine, Department of Human Anatomy, Tikrit, Iraq

***Corresponding author:**

Mohammad Ahmad ABDALLA, Tikrit University - College of Medicine, Department of Human Anatomy, Tikrit, Iraq

E-mail: dr.mohammad68@tu.edu.iq

INTRODUCTION

The thyroid gland was first described by Vesalius, who called it “Glandular laryngitis” since he incorrectly expected that this gland lubricated the larynx. The word thyroid, coined by Thomas Wharton, derived from the Greek word “Thyreos” meaning “an oblong shield” and “Eidos” meaning form¹.

The thyroid gland is a butterfly-shaped gland located just inferior to the larynx (voice box), with the right and left lateral lobes located on either side of the trachea. Connecting the lobes is a mass of tissue called an isthmus that lies anterior to the trachea. The thyroid glands have a small third lobe called the pyramidal lobe². The gland has an abundant blood supply, receiving 80–120 ml of blood per minute³.

Thyroid disorder is a public health problem among a large number of endocrinopathies. About 5% of the world's population is affected by different thyroid diseases. Most of the diseases affecting the thyroid gland, like goiter, are usually associated with enlargement of the gland and require medical and surgical intervention⁴.

The thyroid hormones, thyroxine (T₄), and triiodothyronine (T₃) are essential for normal growth and development⁵. Thyroid-stimulating hormones (TSH) act upon the thyroid gland to initiate thyroid hormone production⁶. Thyroid hormone is necessary for normal brain development; extremely low levels of thyroid hormone during gestation and the neonatal period result in mental retardation⁷. The risk of women compared to men is more elevated (approximately 6/1)⁸. Autoimmune thyroid diseases (AITD) are the most frequent autoimmune disorders and result from an immune assault on the thyroid, and they are classified as “T cell-mediated organ-specific autoimmune disorders”⁹. AITD includes two main clinical presentations: Autoimmune thyroiditis (AT) and Graves' disease (GD). The clinical hallmarks of GD and AT are thyrotoxicosis and hypothyroidism, respectively. The prevalence of AITD is about 5%¹⁰; however, the prevalence of antithyroid antibodies (ATA) may be even higher. The risk of women compared to men is more elevated (approximately 6/1)¹¹. Hypothyroidism is more frequent in older people¹². There is a great geographic variability, and the prevalence of AITD and thyroid antibodies differs with race¹³.

The incidence of AITD is more elevated in subjects living in iodine-sufficient areas in comparison to those in iodine-deficient areas¹⁴. The incidence rate of

AITD is increasing, even if it is not possible to know whether this is due to a higher accuracy of diagnostic procedures¹⁵. The main objective of this study is to evaluate the immunomodulatory role of several factors in patients with thyroid disorders, and then compare them with a control group or healthy subjects. Identify the reasons and know the important future readings of these standards. The objectives of this study were achieved through the following protocol steps: patients with thyroid disorders are diagnosed by a specialist physician, and cytokine concentrations (interleukin 6 (IL-6), interleukin 8 (IL-8), and tumor necrosis factor- α (TNF- α)), are measured using ELISA technology¹⁶.

PATIENTS AND METHODS

The study was carried out at Diyala Teaching Hospital in Baquba, Iraq; from September 2024 to November 2025. The study included 80 female patients, after their condition was confirmed through medical and clinical examinations by doctors who were specialized in this field. Also, a random group that included 20 healthy women was chosen as a control group. The ethical approval was done as a normal process for all human individuals involved in it. *The Medical Ethics Committee of Tikrit University College of Medicine had approved the present study with a code number (IQ.TUCOM.REC.2024.248)*. An ethical standard agreement statement for each individual who participated in the current study, considering the Helsinki Declaration through the World Medical Association, was revised in 2000 at Edinburgh. The management reviewed the study protocol to ensure that it respected participants' rights, maintained confidentiality, minimized potential risks, and upheld principles of beneficence and justice.

The Study design

The study design included grouping patients according to cases of thyroid disorders. The first group included patients with nodular hyperplasia, the second group those who suffered from thyrotoxicosis, the third group those who suffered from hypothyroidism, and the last group those who suffered from papillary thyroid carcinoma.

Collection of Samples

The current study included 80 blood samples collected from females with thyroid disorders and from healthy females. Venous samples were taken before the thyroidectomy was performed for the patients and the control group. Blood samples were drawn from the median

cubital vein or from another vein if it was not accessible. Then 5 ml of blood was taken and placed in the gel tube, and the gel tube was placed in a cooler box (containing an ice pack) until it was transported to the laboratory unit and centrifuged for 10 minutes at 6000 rpm. After whole blood apheresis, the serum was extracted using a micropipette, and then 2 ml of serum was placed in three Eppendorf tubes for immunoassays. They are then stored in a deep freezer (-20°C).

Immunological tests

The levels of Immunological tests [interleukin 6 (IL-6), interleukin 8 (IL-8), and tumor necrosis factor- α (TNF- α)] were estimated by following the steps provided with their ready-made analysis kit according to the manufacturer's special instructions (SunLong Biotech / China) regarding ELISA technology.

Statistical analysis

The statistical analysis was carried out by using a statistical program (Minitab) and comparison between groups, which was made by using one-way analysis of variance ANOVA and testing the arithmetic means for parameters by using Duncan's multiple range test to determine significant difference, especially between groups. The level of statistical significance was taken at ($P \leq 0.01$)¹⁷.

RESULTS AND DISCUSSION

Demographic distribution of the study population

The present study showed that in 80 patients suffering from various thyroid diseases, the final clinical diagnosis was determined as follows: There were 47 cases (59%) suffering from the non-toxic nodular goiter, whereas 11 individuals (14%) suffered from the thyrotoxicosis, and there were 17 individuals (21%) suffering from the hypothyroidism. The findings also revealed that there were 5 individuals suffering from the malignant tumor (6%), as shown in Table 1.

Concentrations of IL-6 in patients and control groups

The (mean $\pm$ SD) of IL-6 concentrations for non-toxic nodular goiter, toxic goiter patients, hypothyroidism, thyroid cancer papillary and control groups respectively were (17.62 ± 1.39) pg/mL, (29.18 ± 3.58) pg/mL, (35.26 ± 2.29) pg/mL, (56.24 ± 5.84) pg/mL, and (8.94 ± 0.97) pg/mL, as shown in Figure 1.

Table 1. Percentage of pathological diagnoses for patients with thyroid gland distributed by number.

Pathological diagnosis	Number	Percentage
G1- Non-Toxic Goiter	47	59 %
G2- Toxic Goiter	11	14 %
G3- Hypothyroidism		
a. Hashimoto	10	21 %
b. Lymphocyte	7	
G4- Thyroid Cancer		
a. Papillary	5	6 %
Total	80	100

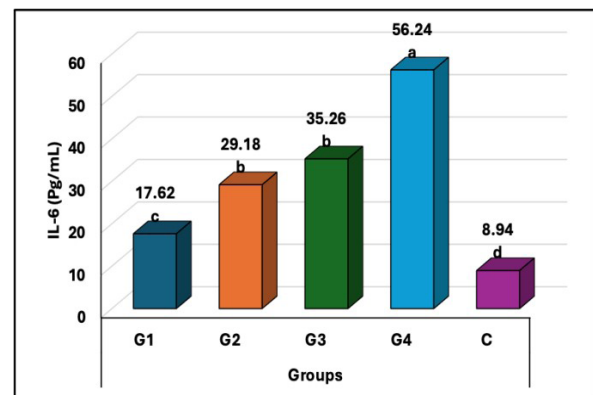


Fig.1. Concentrations of IL-6 (pg/mL) in patients and control groups (*) The similar letters mean that there are no significant differences between groups and the different letters mean that there are significant differences between them at a potential level $P \leq 0.01$ (**). G1:Non-Toxic Goiter, G2:Toxic Goiter, G3:Hypothyroidism, G4:Thyroid Cancer, and C: Healthy Individuals (Control).

A growing body of evidence suggests that inflammation plays a major role in various thyroid diseases. In this study, a significant difference was observed between the five groups (for IL-6); it increased more in the thyroid groups than in the control group.

The significant elevation of IL-6 levels in patients with hyperthyroidism compared to the control group is consistent with studies by Jha et al and Rija et al^{18,19} who focused on this cytokine and reported its important role in the development of hyperthyroidism. They observed that IL-6 plays a role in vascular inflammation because it has different biological activities in different target cells.

The thyroid gland produces IL-6, which is stimulated by antibodies to TSH receptors, IL-1, and possibly TSH receptors. It has been suggested that in patients with GD, the degree of increase may represent the severity of the condition due to higher IL-6 levels

in some studies²⁰. According to a study conducted by Jha et al¹⁸, hyperthyroid patients with active GD ophthalmopathy were found to have higher IL-6 serum concentrations than those without eye disease. Previous studies have shown that hyperthyroidism, especially caused by GD and toxic adenoma, is associated with high levels of IL-6 in the blood. These elevations were not consistent, and it remained difficult to distinguish between the role of thyroid hormones in this elevation and the role of the predominant autoimmune inflammatory process in GD²⁰.

The present results supported the hypothesis that hypothyroidism is associated with increased levels of inflammatory markers compared to controls, which may underlie the risk of cardiovascular disease in patients with hypothyroidism. Found that serum levels of IL-6 were significantly higher in hypothyroid patients compared to euthyroid controls. This is in agreement with Hussein et al., who reported that patients with hypothyroidism had low-grade chronic inflammation, which was confirmed by significant elevation of C-reactive protein (CRP) and IL-6 sensitivity²¹.

The significantly higher concentrations of IL-6 and TNF- α in subclinical hypothyroidism patients were supported by a previous study²². In contrast to the present findings, the hyperthyroid patients had elevated serum levels of the cytokine IL-6, and hypothyroid patients had low levels of IL-6²⁰. According to a study conducted by Shiha et al²³, it was found that increased levels of IL-6 were found in women with hyperthyroidism. However, hypothyroidism did not significantly reduce serum IL-6. Serum IL-6 that was normalized after disease remission suggests that it is related to follicular cell damage.

It found a significant positive relationship between blood TSH levels and inflammatory markers in hypothyroid patients. Gupta²⁴ also reported a significant increase in IL-6 levels in patients with subclinical hypothyroidism compared to healthy thyroid controls, which was also positively associated with serum TSH levels. Abdalla²² found a significant negative correlation between serum total T3 and IL-6, and total T4 and IL-6.

TSH stimulates the release of IL-6 in adipocytes²⁵. IL-6 plays a major role in the early stages of inflammation and also plays a modulatory role during the atherosclerotic process. It helps in accelerating atherosclerosis by producing CRP in the liver²³. TNF- α is involved in systemic inflammation and also induces the acute phase reaction. It is also an important stimulator of IL-6 production²⁶. It acts at every level of the

hypothalamic-pituitary-thyroid axis to produce different changes in the axis²⁷.

These findings were supported by Tayde²⁸ IL-6 was significantly elevated in hypothyroid cases compared to healthy controls, and after treatment with levothyroxine, all markers decreased significantly. Also reported a significantly higher level of TNF- α in patients with hypothyroidism, but did not return to normal after normalization of thyroid function. In contrast, the current study population was newly diagnosed hypothyroid patients, who regained elevated inflammatory markers after treatment with levothyroxine, demonstrating the role of early treatment in resuming normal levels of markers²⁹.

It reported significantly higher levels of serum IL-6 in patients with benign thyroid diseases and thyroid cancer, compared to healthy controls³⁰. Multiple studies have consistently reported that, compared to healthy controls, circulatory IL-6 levels were significantly higher in patients with various conditions such as malignant tumors, colorectal cancer³¹, breast cancer³², pancreatic cancer³³, gastric cancer³⁴, stomach and intestinal cancers³⁵, and others. This study also observed that IL-6 was able to promote the spread of oropharyngeal cancer³⁶.

Concentrations of IL-8 in patients and control groups

The (mean $\pm$ SD) of IL-8 concentrations for non-toxic nodular goiter, toxic goiter patients, hypothyroidism, thyroid cancer papillary and control groups respectively were (16.53 $\pm$ 1.18) pg/mL, (20.52 $\pm$ 2.45) pg/mL, (42.01 $\pm$ 3.62) pg/mL, (61.72 $\pm$ 6.89) pg/mL, and (9.36 $\pm$ 1.27) pg/mL, as shown in Figure 2.

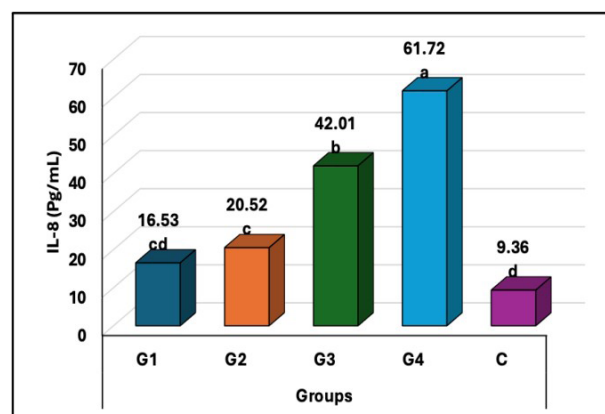


Fig. 2. Concentrations of IL-8 (pg/mL) in patients and control groups. The similar letters mean that there are no significant differences between groups and the different letters mean that there are significant differences between them at a potential level $P \leq 0.01$ (**). G1:Non-Toxic Goiter, G2:Toxic Goiter, G3:Hypothyroidism, G4:Thyroid Cancer, and C: Healthy Individuals (Control).

Serum IL-8 levels were significantly elevated in all patients with thyroid disorders (goiter, autoimmune thyroid disorders, and thyroid cancer) compared to healthy individuals. Statistically significantly higher levels of each of these cytokines were found in patients with PTC. Similar to the present study, it demonstrated significant differences in IL-8 levels in euthyroid patients and normal controls³⁷.

Also, significantly elevated levels of IL-8 in patients with GD and nodular goiter compared to respective healthy control groups³⁸. In contrast to these studies, Krassas and colleagues found that IL-8 levels were not elevated in GD, toxic nodular goiter, and HT³⁹. Another study showed increased levels of cytokines, including IL-8, in patients who repeatedly develop thyroid disease syndromes followed by allogeneic bone marrow transplantation⁴⁰.

Increased levels of IL-8 have also been observed in patients with other malignant tumors. It was also found that the serum level of IL-8 was significantly elevated in most patients with Hepatocellular Carcinoma (HCC) compared with healthy subjects⁴¹. In fact, an elevated serum level of IL-8 has been found to be a diagnostic marker in soft tissue sarcoma⁴², B-cell chronic lymphocytic leukemia⁴³, primary gastrointestinal non-Hodgkin lymphoma⁴⁴, and malignant melanoma⁴⁵. Therefore, the elevated levels of both cytokines in the serum of thyroid cancer patients may be due to their overproduction in cancer cells and their subsequent release into the circulation. The incidence of thyroid cancer in thyroid cancer patients with higher levels of IL-8 compared to healthy subjects increased with increasing disease stage.

High serum IL-8 level correlates with large tumor size and advanced tumor stage in patients with HCC⁴⁶. Increased IL-8 expression has been found in many tumors, and in some studies, IL-8 serum and/or tissue levels correlate with tumor progression and metastasis⁴⁷. Therefore, this finding also supports a role for IL-8 in thyroid cancer development. Finally, monitoring serum IL-8 levels can help differentiate patients with thyroid disease from healthy individuals, and IL-8 appears to play a role in the pathogenesis of thyroid disease and may represent a target for innovative diagnosis and treatment.

The difficulties encountered likely arise during the differential detection of benign and malignant thyroid conditions because they both have commonalities in basic mechanisms. It is well documented that hypothyroidism cancer often occurs in the context

of autoimmune thyroid diseases (AITDs), especially Hashimoto's thyroiditis (HT) and Graves' disease (GD)⁴⁸. It has already been suggested that AITDs predispose to malignant transformation by establishing the tumorigenic microenvironment, where RET/PTC rearrangements are well tolerated to maintain inflammation operations⁴⁹. In fact, many studies have documented that it alters the production of cytokines and chemokines in GD and HT, resulting in the initiation and maintenance of inflammatory and immune reactions⁵⁰.

In this study, a large percentage of patients suffer from thyroid disease, and they were also diagnosed with thyroiditis, primarily HT. And so on, activation of inflammation-related mechanisms, in particular induction of the inflammatory transcription program, was observed in both benign and malignant thyroid glands. The conditions make their differentiation by cytokine concentrations somewhat difficult, and provide a reasonable solution and explanation for the lack of statistically significant differences between patient groups in our observations.

Concentrations of TNF- α in patients and control groups

The (mean $\pm$ SD) of TNF- α concentrations for non-toxic nodular goiter, toxic goiter patients, hypothyroidism, thyroid cancer papillary and control groups respectively were (15.66 $\pm$ 0.92) pg/mL, (27.35 $\pm$ 2.85) pg/mL, (27.10 $\pm$ 2.46) pg/mL, (56.89 $\pm$ 5.03) pg/mL, and (12.37 $\pm$ 0.99) pg/mL, as shown in Figure 3.

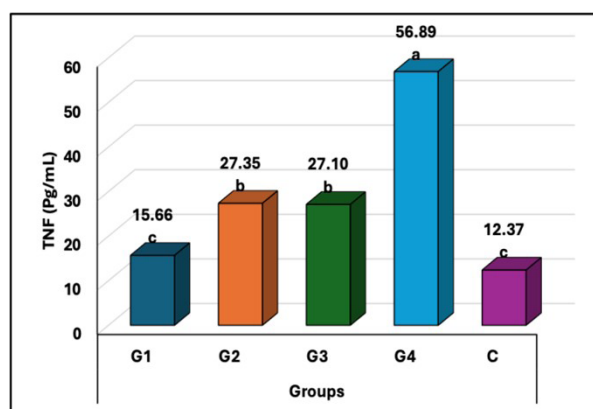


Fig. 3. Concentrations of TNF- α (Pg/mL) in patients and control groups. The similar letters mean that there are no significant differences between groups and the different letters mean that there are significant differences between them at a potential level $P \leq 0.01^{**}$. G1: Non-Toxic Goiter, G2: Toxic Goiter, G3: Hypothyroidism, G4: Thyroid Cancer, and C: Healthy Individuals (Control).

Serum TNF- α levels were significantly elevated in patients with thyroid disorders (PTC, GD, and HT) compared to healthy individuals, whereas there were no significant differences in patients with goiter. TNF- α : It is an important cytokine involved in systemic inflammation⁵¹. TNF- α is an inflammatory cytokine produced mainly by monocytes and macrophages with pleiotropic effects. It binds to cell surface receptors and prevents the usual rise in intracellular calcium concentrations, resulting in the activation of multiple signal transduction pathways, enzymes, and transcription factors⁵².

The present study revealed that serum TNF- α was significantly higher in PTC patients, compared to healthy individuals. However, it noted that patients with thyroid disease tend to have lower TNF- α level compared to normal controls⁵³, and TNF- α act as a growth inhibitor of PTC cell lines⁵⁴. Another study showed that the average TNF- α levels measured in tumor patients were not significantly different from those observed in the control group⁵⁵.

Cytoplasmic TNF- α was found to be significantly higher in primary tumors of PTC patients compared to patients with benign thyroid diseases. TNF- α expression was only significantly positively associated with the presence of calcification in tumors, suggesting that TNF- α is likely involved in the development of calcification in tumors⁵⁶. The present results may use TNF- α as a risk indicator for PTC and may have a role in tumor progression and prognosis in patients with thyroid cancer. Investigation into the role of cytokines in the onset and course of autoimmune thyroid diseases has been ongoing for decades. The pro-inflammatory cytokine, TNF- α , is involved in the induction and maintenance of immune responses. It is thought to play a crucial role in autoimmune thyroid diseases, including GD and HT⁵⁷.

Since it has been reported that TNF- α is usually short-lived in peripheral blood, various changes in TNF- α serum levels have been reported in patients with hyperthyroidism and hypothyroidism. In contrast, High serum levels of TNF- α have been reported in patients with GD, who were unable to return to normal after 6 weeks of taking antithyroid medications. Elevated levels of TNF- α in the serum of GD patients may affect the systemic response to the disease or the reciprocity between antigen and antibody within the thyroid gland. Increased cytokines in the thyroid may lead to the production of other cytokines by autoreactive cells, thus perpetuating the autoimmune reaction⁵⁸.

CONCLUSION

The results of this study indicate that all types of thyroid disorders are more common in females than in males. It was noted that thyroid cancer was less common among thyroid diseases, as pathological changes were observed, in addition to noticeable alterations in the levels of cytokines. The incidence of thyroid diseases is not limited to a specific demographic area, and the study results support the fact that cytokines play essential roles in the development and prognosis of thyroid disorders. The current study indicates that measurements (TNF- α , IL-8, and IL-6) can be considered diagnostic tests to predict thyroid disorders.

Ethics Statement and Conflict of Interest Disclosures

Financial support and sponsorship: All authors have declared that no financial support was received from any organization for the submitted work.

Ethics Consideration: The authors declare that all the procedures and experiments of this study respect the ethical standards in the Helsinki Declaration of 1975, as revised in 2008(5), as well as the national laws. Written informed consent was provided by the patient participant in this study. This study was approved by the Institutional Research Board and Ethics Committee.

Conflict of interest: No known conflict of interest correlated with this publication.

Availability of data and materials: The data used and/ or analyzed throughout this study are available from the corresponding authors upon reasonable request.

Competing interests: The authors declared that they have no competing interests.

The use of generative AI and AI-assisted technologies: The authors did not use in this article generative AI and AI-assisted technologies.

References

1. Lorrence A. Thyroid gland: small but mighty. *North Light Ear Head Throat Clin.* 2006;715:735-8.
2. Tortora GJ, Nielsen M. Principles of human anatomy. Hoboken (NJ): John Wiley & Sons; 2017; pp. 312-451.
3. Li Y, Wang X, Chen H, Yan L, Li M, Zhang M et al. Ultrasound-guided microwave ablation combined with ethanol injection for the treatment of solitary nodular retrosternal goiter: a prospective study of 72 patients. *Eur Radiol.* 2023;33(2):752-62.

4. Chandra AK. Mechanism of development of residual goitre and associated thyroid disorders. *Indian J Physiol Allied Sci.* 2022;74(4):24-36.
5. Forhead AJ, Fowden AL. Thyroid hormones in fetal growth and prepartum maturation. *J Endocrinol.* 2024;221(3):R87-103.
6. Mullur R, Liu YY, Brent GA. Thyroid hormone regulation of metabolism. *Physiol Rev.* 2014;94(2):355-82.
7. Fini JB, Le Mével S, Turque N, Leemans M, Lettmann M, Spirhanzlova P et al. Human amniotic fluid contaminants alter thyroid hormone signalling and early brain development in *Xenopus* embryos. *Sci Rep.* 2017;7(1):43786.
8. Rizzo M, Pernice V, Franchina T. Increased annual frequency of Hashimoto's thyroiditis between years 1988 and 2007 at a cytological unit of Sicily. *Ann Endocrinol (Paris).* 2024;71(6):525-31.
9. Zhang Y, Sun H. The emerging function and clinical significance of circRNAs in thyroid cancer and autoimmune thyroid diseases. *Int J Biol Sci.* 2023;17(7):1731-45.
10. Ovčariček PP, Görges R, Giovannella L. Autoimmune thyroid diseases. *Semin Nucl Med.* 2023;53(2):128-39.
11. Patti M, Christian R, Palokas M. Association between anti-thyroid antibodies and quality of life in patients with Hashimoto thyroiditis: a systematic review and meta-analysis. *JBI Evid Synth.* 2021;19(9):2307-38.
12. de Montmollin M. L-thyroxine therapy for older adults with subclinical hypothyroidism and hypothyroid symptoms: secondary analysis of a randomized trial. *Ann Intern Med.* 2024;172(11):709-16.
13. Conigliaro P. Autoimmune thyroid disorders and rheumatoid arthritis: a bidirectional interplay. *Autoimmun Rev.* 2023;19(6):102529.
14. Wan S, Franchina T. Autoimmune thyroid diseases after 25 years of universal salt iodisation: an epidemiological study of Chinese adults in areas with different water iodine levels. *Br J Nutr.* 2022;124(8):853-64.
15. Zhang X. Prevalence of autoimmune thyroid disease in patients with psoriasis: a meta-analysis. *BMJ Open.* 2022;12(1):e055538.
16. Antonelli A. Autoimmune thyroid disorders. *Autoimmun Rev.* 2025;14(2):174-80.
17. Popović BV. Understanding advanced statistical methods. *J Applied Statistics.* 2014; 41(12): 2777 -87.
18. Jha RK, Kondhalkar AA, Kute R. Level of interleukin 6, malondialdehyde and calcium in hypo and hyperthyroidism. *Eur J Mol Clin Med.* 2024;8(1):208.
19. Rija FF, Hussein SZ, Abdalla MA. Osteoprotegerin, sclerostin, and osteocalcin serum levels in thyroid disorder patients. *Ukr Biochem J.* 2021;93(5):117-21. doi:10.15407/ubj93.05.117
20. Abdalla MA, Nayaf MS, Hussein SZ. Correlation between serum α -MSH and vitamin D levels in vitiligo patients. *Iran J Dermatol.* 2020;23(4):163-7. doi:10.22034/ijd.2020.120836
21. Hussein SZ, Abdalla MA. Serum levels of alpha-melanocyte stimulating hormone, vitamin D, calcium, phosphorus and magnesium in COVID-19 patients. *Ukr Biochem J.* 2021;93(6):64-9. doi:10.15407/ubj93.06.064
22. Abdalla MA. The prevalence of pyramidal lobe of the thyroid gland among Iraqi society. *Tikrit Med J.* 2016;21(1):135-40.
23. Shihan JT, Al-Janabi MSA, Abdalla MA. Impact of aqueous extract of ginger and chamomile plant and some antihistamines in the treatment of drug-allergy induced in white rats: a physiological and immunological study. *HIV Nurs.* 2022;22(2):3449-54.
24. Gupta G. Study on subclinical hypothyroidism and its association with various inflammatory markers. *J Clin Diagn Res.* 2025;9(11):BC04.
25. Antunes TT. Thyroid-stimulating hormone stimulates interleukin-6 release from 3T3-L1 adipocytes through a cAMP-protein kinase A pathway. *Obes Res.* 2005;13(12):2066-71.
26. Wang T, He C. TNF- α and IL-6: the link between immune and bone system. *Curr Drug Targets.* 2020;21(3):213-27.
27. Abdalla MA. The vitiligo is the human skin's pigmentary challenge still: an up-to-date review. *Probl Sotsialnoi Gig Zdravookhranennii Istor Med.* 2024;32(5):985-96. doi:10.32687/0869-866X-2024-32-5-985-996
28. Tayde PS. Hypothyroidism and depression: are cytokines the link? *Indian J Endocrinol Metab.* 2023;21(6):886-92.
29. Qasim YA, Salih AA, Mahmood EM, Abdalla MA. Histological study of coronary arteries in different ages in Kirkuk governorate. *Azerbaijan Med J.* 2022;62(6):2169-75.
30. Zhang L. Inflammatory tumor microenvironment of thyroid cancer promotes cellular dedifferentiation and silencing of iodide-handling genes expression. *Pathol Res Pract.* 2023;246:154495.
31. Hussein SZ, Abdalla MA. Levels of leptin, adiponectin, and insulin in COVID-19 patients. *Medicina Moderna.* 2022;29(1):89-92. doi:10.31689/rmm.2021.29.1.89
32. Tripsianis G, Papadopoulou E, Anagnostopoulos K. Coexpression of IL-6 and TNF- α : prognostic significance on breast cancer outcome. *Neoplasma.* 2014;61(2):205-12.
33. Abdalla MA, Nayaf MS, Hussein SZ. Evaluation of vitamin D in melasma patients. *Rev Rom Med Lab.* 2019;27(2):219-21. doi:10.2478/rrlm-2019-0023
34. Krzystek-Korpacka M. Impact of weight loss on circulating IL-1, IL-6, IL-8, TNF- α , VEGF-A, VEGF-C and midkine in gastroesophageal cancer patients. *Clin Biochem.* 2023;40(18):1353-60.
35. Kobawala TP. Significance of interleukin-6 in papillary thyroid carcinoma. *J Thyroid Res.* 2024; 16:1-8.
36. Rija FF, Hussein SZ, Abdalla MA. Physiological and immunological disturbance in rheumatoid arthritis patients. *Baghdad Sci J.* 2021;18(2):247-52. doi:10.21123/bsj.2021.18.2.0247
37. Siddiq A. Association of pro-inflammatory cytokines with vitamin D in Hashimoto's thyroid autoimmune disease. *Medicina.* 2023;59(5):853.
38. Antonelli A. Graves' disease: clinical manifestations, immune pathogenesis (cytokines and chemokines) and therapy. *Best Pract Res Clin Endocrinol Metab.* 2020;34(1):101388.

39. Krassas GE, Bougoulia M, Koliakos G. Serum interleukin-8 levels in thyroid diseases. *Thyroid*. 2019;10(5):445-6.
40. Rija FF, Hussein SZ, Abdalla MA. Study the effect of some adipokines and interleukins in hypo and hyperthyroidism patients. *Medicina Moderna*. 2023;30(3):199-203. doi:10.31689/rmm.2023.30.3.199
41. Zhong M, Zhang L. Serum interleukin-8 as an indicator of tumor recurrence in hepatocellular carcinoma after transarterial chemoembolization. *Int J Clin Exp Med*. 2024;13(5):3540-7.
42. Koseci T. Prognostic importance of inflammatory indexes in patients treated by pazopanib for soft tissue sarcoma. *Clin Lab*. 2022;68(2):1-7.
43. Abdalla MA. Melasma clinical features, diagnosis, epidemiology and etiology: an update review. *Siriraj Med J*. 2021;73:841-50. doi:10.33192/Smj.2021.109
44. Retzlaff S, Turque N, Leemans M. Interleukin 8 and Flt3 ligand as markers of advanced disease in primary gastrointestinal non-Hodgkin's lymphoma. *Oncol Rep*. 2022;9(3):525-7.
45. Ugurel S. Increased serum concentration of angiogenic factors in malignant melanoma patients correlates with tumor progression and survival. *J Clin Oncol*. 2021;19(2):577-83.
46. Öcal O. Baseline interleukin-6 and -8 predict response and survival in patients with advanced hepatocellular carcinoma treated with sorafenib monotherapy: an exploratory post hoc analysis of the SORAMIC trial. *J Cancer Res Clin Oncol*. 2022;148(9):2411-22.
47. Abdalla MA, Nayaf MS. Evaluation of serum α -MSH level in melasma. *WJPMR*. 2018;4(5):29-32.
48. Casto C. Hashimoto's thyroiditis and Graves' disease in genetic syndromes in pediatric age. *Genes (Basel)*. 2023;12(2):222.
49. Murthy SP, Vidhyadharan S, Subramaniam N. Thyroid malignancies. *Comprehensive management of head and neck cancer*. 2021;1:299.
50. Gallo D. How does vitamin D affect immune cells crosstalk in autoimmune diseases? *Int J Mol Sci*. 2023;24(5):4689.
51. Mastrangelo F. New concepts in neuroinflammation: mast cells pro-inflammatory and anti-inflammatory cytokine mediators. *J Biol Regul Homeost Agents*. 2018;32(2):449-54.
52. Wu CK, Chen H, Yan L, Li M. Plasma levels of tumor necrosis factor- α and interleukin-6 are associated with diastolic heart failure through downregulation of sarcoplasmic reticulum Ca^{2+} ATPase. *Crit Care Med*. 2011;39(5):984-92.
53. Hodzic-Redzic S. The role of preoperative levels of serum IL-6, IL-8 and TNF- α and conventional inflammatory parameters in the detection of metastatic forms of papillary thyroid cancer. *WCRJ*. 2021;8:e1915.
54. Lumachi F, Basso SMM, Orlando R. Cytokines, thyroid diseases and thyroid cancer. *Cytokine*. 2010;50(3):229-33.
55. Gendek-Kubiak H. Serum TNF-alpha level in neoplasm patients qualified for surgical treatment. *Arch Immunol Ther Exp (Warsz)*. 2001;49(Suppl):S97-102.
56. Wu CK, Li M. Plasma levels of tumor necrosis factor-alpha and interleukin-6 are associated with diastolic heart failure through downregulation of sarcoplasmic reticulum Ca^{2+} ATPase. *Crit Care Med*. 2011;39(5):984-92.
57. Li W, Chen H. TRIM30 modulates interleukin-22-regulated papillary thyroid cancer cell migration and invasion by targeting Sox17 for K48-linked polyubiquitination. *Cell Commun Signal*. 2019;17(1):1-14.
58. Çelik I, Akalin S, Erbaş T. Serum levels of interleukin 6 and tumor necrosis factor- α in hyperthyroid patients before and after propylthiouracil treatment. *Eur J Endocrinol*. 1995;132(6):668-72.